

Study on the Applicability of Barcode Loci at the Generic Level among Congeneric Species

Komal^{1*} Dr. Krishan Pal Singh²

¹ Research Scholar of OPJS University, Churu, Rajasthan

² Associate Professor, OPJS University, Churu, Rajasthan

Abstract – *Debasement in the medicinal plant material is the common factor presenting negative consequences for the commercialization of medicinal plants. Both purposeful and unplanned blending and substitution of unique medication crude material with the less successful species lessens the nature of the last items. Utilization of credible and unique species is the absolute first and basic advance to deliver great quality, dependable and effectual home grown medications. Logical legitimacy of the species is the first and fundamental parameter for quality confirmation of the plant based therapeutics. Numerous customary procedures like macroscopy, microscopy and detachment of synthetic marker mixes have been utilizing since long to recognize bona fide plants and to isolate them from their adulterants. Genome based distinguishing proof strategies give an elective device which can be utilized freely or synergistically with the regular techniques. Amalgamation of DNA based methods with the customary strategies of distinguishing proof present a viable methodology for quality control and affirmation of medicinal plants.*

Keywords – Barcode, DNA, Congeneric Species

-----X-----

INTRODUCTION

DNA barcoding is a technique, which gives speedy recognizable proof of species without including the morphological prompts. It utilizes a generally little institutionalized DNA part as a tag, to characterize or find a species. DNA barcoding utilizes minor contrasts of nucleotides in the specific quality loci of various life forms as a key for the segregation. The quality is sequenced to know the base-pair contrasts and afterward stored in the barcode database, which is named as DNA barcodes. These hereditary codes could be gotten to through a computerized library and used to recognize the obscure species by any researcher around the globe. Perfect DNA barcode ought to be regularly a uniform short arrangement of DNA (400-800 bp), ready to be essentially produced and used to describe all the living beings (Savolainen et al., 2005). Paul's gathering was the first to plan and utilize the short DNA groupings for natural ID at the University of Guelph, Canada. The possibility of barcoding was first developed to portray the microorganisms, in which the morphological keys were inadequate. Presently it is being connected effectively to creatures. A gigantic on-line advanced library of barcodes will be a standard, to which the DNA barcode arrangement of an obscure example can be coordinated for the distinguishing proof. The key procedure in DNA barcoding is distinguishing novel applicant quality, which can be utilized all around. It ought to permit the clients proficiently to recognize the

species and quicken the species revelation. DNA barcoding utilizes the data of one or a couple of districts in the genome to perceive every one of the species in a family (Lahaye et al., 2008). DNA barcoding will open up new open doors in DNA based examinations going from network phylogenetics (Webb et al., 2002) to environmental genomics (van Straalen and Roelofs, 2006). In spite of the absence of an all-inclusive plant barcode, taxonomists, environmentalists, transformative researcher and protectionists are for the most part previously imagining the motivation behind a hereditary identifier to a wide arrangement of research and viable applications. The appropriateness of a locus for enormous scale DNA barcoding can be effectively examined by contrasting loci over the comparable arrangement of taxa under a chose set of PCR conditions.

In this way, the measurements was considered between the capacity to enhance a locus and the pace of uniqueness of that locus over a phylogenetic scope of taxa. Furthermore, the succession arrangement strategies are accessible, which can be assessed for the utilization of DNA barcodes with the accompanying respects,

- The motivation behind affirmation breaking points to species task

- The utilization of a piece of groupings in database look
- The quality of pursuit calculations of grouping length variety because of inclusion/erasure occasions and the enlightening idea of these transformations.

DNA bar-coding mirrors the appropriation of intra and intra-explicit variety that is isolated by a separation called „DNA barcoding gap“ (Hebert et al., 2003, Meyer and Paulay, 2005). The Consortium of Barcode of Life organizes DNA barcoding advancement and usage generally. DNA barcoding is extremely basic for the sub-atomic ID of effectively depicted species (Hebert et al., 2003) and the revelation of new species (Valentini et al., 2006).

REVIEW OF LITERATURE

The family Orchidaceae is one of the exceptionally advanced and biggest groups of angiosperms with around 25,000-35,000 species having a place with 750 to 850 genera appropriated around the world (Chase 2005, Hossain 2011). This profoundly propelled group of monocots is spoken to by for the most part herbaceous plants which are epiphytes, lithophytes, saprophytes or earthly; have particular flower morphology, fertilization instruments and moment seeds; and are related with remarkable parasitic accomplices (mycorrhizae), (Kumar et al. 2007). The trademark includes by which orchids can be recognized from different plants are: respective symmetry/zygomorphic blossoms, nearness of labellum or lip (profoundly changed petal), combined stamens and carpels shaping particular structure called gynostemium or section, mediocre ovary which is commonly turned to 180° (resupinate blooms) and amazingly moment seeds (Chowdhery 1998).

Orchids are conveyed worldwide mostly in the wet tropics. They have not been accounted for from the polar district and the driest of deserts (Chase 2005). The higher-level characterization of Orchidaceae has generally been founded on the development of gynostemium or section. The segment in light of its subtleties is one of a kind to the family (Chase 2005). The family Orchidaceae contains a few megagenera with 1000+ sp. e.g., *Bulbophyllum*, *Epidendrum*, *Pleurothallis* and *Dendrobium* (Whitten et al. 2007). The explanations behind such dangerous speciation, perhaps on account of natural adjustments, physiological/morphological developments, or quickened paces of morphological/sub-atomic change, are as yet not appropriately comprehended (Whitten et al. 2007).

In any case, orchid species are commonly hard to recognize as these are grouped fundamentally based on botanical morphology that changes with pollinator inclinations (Cameron 2004). In this manner, for better circumscription and inducing connections among the hard to characterize Orchidaceae genera, various

gatherings, using DNA districts from the mitochondrial, plastid and atomic genomes, have embraced sub-atomic precise investigations. Be that as it may, before examining the atomic systematics of orchids, a short evaluation of the morphology-based characterization of the family is fundamental.

Dressler (1993) proposed the most recent and most acknowledged characterization of the Orchidaceae that was in view of on the anther morphology. As per this grouping, the family includes 850 genera and 20,000 species. These were organized in 70 sub-clans, 22 clans and five sub-families (Dressler 1993).

Notwithstanding none-too-attractive circumscription of the Neottioideae, the other real gathering of orchids that was dangerous was Vanilloideae, the sections of which resemble those of the Epidendroideae, yet vegetatively they are exceptionally dissimilar from every single other orchid (Cameron and Dickison 1998, Stern and Judd 2000, Chase 2005).

Orchidaceae when contrasted with the morphological cladistics of Dressler (1993), talked about in detail in later area. Ensuing to these two starting reports, various DNA based phylogenetic investigations have been distributed, at the degree of family (Freudenstein et al. 2004, Cameron 2004), sub-families (Neyland and Urbatsch 1995, 1996, Cox et al. 1997, Freudenstein and Chase 2001), clans (Douzery et al. 1999, Kores et al. 2001, Goldman et al. 2001) and sub-clans (Yukawa et al. 1996, Cozzolino et al. 1996, Pridgeon et al. 2001, van nook Berg et al. 2005, Clements 2003).

The information gave proof to the monophyly of at any rate four sub-families. In any case, the degree of arrangement variety found in this quality area was again not adequate for tending to connections underneath the position of sub-family or clan (Freudenstein and Chase 2001).

The powerlessness of atomic and mitochondrial qualities in clarifying the phylogenetic connections underneath sub-family or innate level made the specialists to switch back to plastid qualities. The upsides of utilizing plastid qualities incorporate one-sided legacy, various duplicates per cell, simplicity of intensification and sequencing, just as nonattendance in creatures and organisms (Palmer et al. 1988, Clegg and Zurawski 1992, Olmstead and Palmer 1994). The last one is particularly critical for investigations of Orchidaceae since they normally live in cooperative relationship with contagious accomplices. An extended plastid phylogeny for the family was accounted for by Freudenstein et al. (2004).

Atomic connections of different sub-clans inside Vandaeae were surveyed by the examinations of the information from nucleotide groupings of nrITS, trnL-F, and matK. Greatest stinginess examinations of these three DNA areas upheld just two sub-clans

inside the monopodial Vandeeae. The two sub-clans perceived were the first Aeridinae and Angraecinae, with the third sub-clan Aerangidinae converged in the last mentioned. Independently, Aerangidinae and Angraecinae were observed to be polyphyletic, yet together they framed a well-bolstered monophyletic bunch in every single atomic investigation (Carlsward et al. 2006).

Hidayat et al. (2005) and Micheneau et al. (2008), separately have done the sub-atomic phylogenetic of sub-clans Aeridinae and Angraecinae having a place with the clan Vandeeae. The DNA arrangements from matK and ITS were analyzed from 75 species speaking to 62 genera of sub-clan Aeridinae to find connections (Hidayat et al. 2005).

Salazar et al. (2009) contemplated the phylogenetic connections of sub-clans Cranichidinae and Prescottiinae from the clan Cranchideae, while Álvarez-Molina and Cameron (2009) surveyed phylogenetic relationships among the individuals from just the last sub-clan. Salazar et al. (2009) assessed nucleotide groupings from both atomic ribosomal (ITS) and plastid DNA (rbcL, matK-trnK and trnL-trnF) with cladistic stringency and Bayesian surmising for 45 species having a place with 14 genera of Cranichidinae and Prescottiinae (counting appropriate out-gatherings).

Yukawa et al. (1993) assessed between conventional connections in the sub-clan (Podochileae) at atomic level by utilizing plastid DNA confinement destinations in 15 species having a place with 12 segments of the sub-clan. The outcomes showed that the sub-clan Dendrobieae involved three noteworthy clades.

The twelve Taiwan species of *Dendrobium* were arranged and the hereditary connections were surmised utilizing ITS successions by Tsai et al. (2004). The hereditary separation was determined utilizing Kimura-2-parameter technique and among the 12 *Dendrobium* species, the scope of hereditary separations was from 0.06 to 0.28. Every *Dendrobium* species could be effectively recognized dependent on the ITS arrangement as deduced by hereditary separation and Neighbor Joining techniques.

Numerous orchidaceous species have likewise been utilized in customary arrangement of prescription for restoring different afflictions like tuberculosis, loss of motion, stomach issue, chest torment, joint inflammation, syphilis, jaundice, cholera, acidity, dermatitis, tumor, heaps, bubbles, irritations, menstrual confusion, spermatorrhea, leucoderma, diarrhoea, solid agony, blood looseness of the bowels, hepatitis, dyspepsia, bone breaks, ailment, asthma, intestinal sickness, ear infection, explicitly transmitted illnesses, wounds and injuries (Bulpitt et al. 2007, Hossain 2011). The remedial properties of various orchids are: love potion, rejuvenator, tonic, antibacterial, cancer prevention agent and immunomodulation (Bulpitt et al. 2007).

OBJECTIVES

1. DNA barcode have applications in different fields like, environment, biomedicine, the study of disease transmission, developmental science, biogeography, preservation science and in bio-industry.
2. DNA barcoding must be sensible for a wide scope of experts and the procedure must be available & the quality of DNA barcodes must be straightforward and effective.
3. DNA barcoding to plants had turned out to be suspect in light of the absence of understanding about a center barcode, regardless of whether dependent on a solitary locus or different loci.
4. Plants have not been given much significance in the beginning times of DNA barcoding because of failure of cytochrome oxidase (COX1) to function as a barcode But now it has an immense job in the preservation science.

CONCLUSIONS

The present investigation uncovered ITS to be the best DNA barcode managing most astounding species goals in both the arrays (informational collection I and II). The first contained species from the four of the five sub-families Orchidaceae alongside non-orchid species from different groups of plants and the second was spoken to by just orchids. These outcomes clearly focuses towards appropriateness of ITS as the DNA barcodes for the land plants, as has been recommended as of late by some different agents as well. Be that as it may, this locus alone couldn't be an all-inclusive barcode for plants in light of the few confining reasons talked about in the proposition. Among the loci tried from the chloroplast genome, mat K gave best species goals esteems in the two species arrays. Notwithstanding when connected to congeneric species of twenty orchid genera, these two loci gave best species goals. Or maybe, whenever taken together, these two loci yielded 100% species goals. The other much advertised locus, rbcL, is by all accounts successful at higher ordered level, in this way pointing towards its utility in staggered barcodes.

Among the multi-locus mixes tried for 76 orchid species, the most elevated species goals was achieved by a three-locus mix of ITS+matK+rbcL with 96.05% species goals. The two-locus mix from the chloroplast genome (matK+rbcL), proposed as all inclusive barcode for land plants by CBOL Plant Working Group, gave 90.79% species separation accomplishment among 76 species. This is more than the species goals of 72% gotten by them in a

floristic gathering of species based on which this suggestion was made. Be that as it may, ITS+matK displayed still higher species goals (94.74%) than the past blend. The present examination showed that a three-locus blend (matK+rbcL+ITS), one of the mixes proposed as of late by another gathering as DNA barcode for the land plants, could resolve 96.05% species, yet there was no expansion in species goals when the staying two loci viz., rpoB and rpoC1 from the chloroplast genome were added to the above mix as four-or potentially five-locus mixes. These perceptions show the utility of incorporating ITS in the center DNA barcode of matK+rbcL for orchid species.

The outcomes exhibited in the postulation, enough location to the worries about the appropriateness of DNA barcoding to plants. Be that as it may, journey for an ideal all inclusive barcode for plants giving 100% species goals over the plant kingdom seems, by all accounts, to be unreasonable, as DNA barcoding, similar to some other innovation, isn't required to be 100% flawless. In any case, inside a scientific categorization 100% species goals could be acquired by taxa explicit barcodes. Along these lines, the projection that DNA barcodes, when accessible for all the depicted species, would almost certainly give a right personality up to species level to any obscure example, regardless of whether accessible in vegetative, divided or DNA structure, or would show the disclosure of another species does not remain constant. All things considered, over 90% achievement in species ID with single locus or two-/three-locus mixes earnestly shows the adequacy of the technique. The cases of disappointment of DNA barcodes to effectively allot the species ought to urge taxonomist to re-consider or reinvestigate such taxa.

REFERENCE

1. Adams PB (2011). Systematics of Dendrobieae (Orchidaceae), with special reference to Australian taxa. *Bot J Linn Soc*, 166: pp. 105-126.
2. Alvarez I, Wendel JF (2003). Ribosomal ITS sequences and plant phylogenetic inference. *MolPhylogenetEvol* 2003, 29: pp. 417-434.
3. Álvarez-Molina A, Cameron KM: Molecular phylogenetics of Prescottiinae (2009) and their close allies (Orchidaceae, Cranichideae) inferred from plastid and nuclear ribosomal DNA sequences. *Am J Bot*, 96(5): pp. 1020-1040.
4. Baldwin BG, Sanderson MJ, Porter JM, Wojciechowski MF, Campbell CS, Donoghue MJ (1995). The ITS regions of nuclear ribosomal DNA: a valuable source of evidence on angiosperm phylogeny. *Ann Mo Bot Gard*, 82: pp. 247-277.
5. Barnwell P, Blanchard AN, Bryant JA, Smirnoff N, Weir AF (1998). Isolation of DNA from the highly mucilaginous succulent plant *Sedum telephium*. *Plant MolBiolRepr*, 16: pp. 133-138.
6. Cameron KM, Chase MW, Whitten WM, Kores PJ, Jarrell DC, Albert VA, Yukawa T, Hills HG, Goldman DH (1999). A Phylogenetic analysis of the Orchidaceae: evidence from rbcL nucleotide sequences. *Am J Bot*, 86: pp. 208-224.
7. Delforge P. (1994). "Guide des Orchidées d'Europe, d'Afrique du Nord et du Proche-Orient," Delachaux et Niestlé, Lausanne.
8. Dressler RL (1974). Classification of the orchid family. In: Ospina M (ed.) *Proc 7th World Orchid ConfMedellín*, Colombia pp. 259-274.
9. Eaton MJ, Meyers GL, Kolokotronis SO, Leslie MS, Martin AP, Amato G (2010). Barcoding bushmeat: molecular identification of Central African and South American harvested vertebrates. *Cons Genet*, 11: pp. 1389-1404.
10. Ebihara A, Nitta JH, Ito M (2010). Molecular species identification with rich floristic sampling: DNA barcoding the pteridophyte flora of Japan. *PLoS One*, 5:e15136.
11. Farrington L, Mac Gillivray P, Faast R, Austin A (2009). Investigating DNA barcoding options for the identification of *Caladenia* (Orchidaceae) species. *Aust J Bot*, 57: pp. 276-286.
12. Fazekas AJ, Burgess KS, Kesanakurti PR, Graham SW, Newmaster SG, Husban BC, Percy DM, Hajibabaei M, Barrett SCH (2008). Multiple multilocus DNA barcodes from the plastid genome discriminate plant species equally well. *PLoS One*, 3:e2802.
13. Gao T, Sun Z, Yao H, Song J, Zhu Y, Ma X, Chen S (2011). Identification of Fabaceae plants using the DNA barcode matK. *Planta Med*, 77: pp. 92-94.
14. Garay L. A. (1972). On the origin of the Orchidaceae, II. *J Arnold Arboretum*, 53: pp. 202-215.
15. Govaerts R, Dransfield J, Zona, SF, Hodel DR, Henderson A (2010). World Checklist of Arecaceae. Facilitated by the Royal Botanic Gardens, Kew. <http://apps.kew.org/wcsp/> 2010.

16. Harmon L (2010). Rare flowers and common herbal supplements get unmasked with plant DNA barcoding (reporting unpublished data of Damon Little and David Baker). *Scientific American* Observations, <http://www.scientificamerican.com/blog/post.cfm?id=rare-flowers-and-common-herbalsupp-2010-04-18>.
17. He J, Wong KL, Shaw PC, Wang H, Li DZ (2010). Identification of the medicinal plants in *Aconitum L* by DNA barcoding technique. *Planta Med*, 76: pp. 1622-1628.
18. Kocyan A, de Vogel EF, Elena Conti E, Gravendeel B (2008). Molecular phylogeny of *Aerides* (Orchidaceae) based on one nuclear and two plastid markers: A step forward in understanding the evolution of the *Aeridinae*. *MolPhylogenetEvol*, 48: pp. 422-443.
19. Lahaye R, van der Bank M, Bogarin D, Warner J, Pupulin F, Gigot G, Maurin O, Duthoit S, Barraclough TG, Savolainen V (2008). DNA barcoding the floras of biodiversity hotspots. *Proc Natl AcadSci USA*, 105: pp. 2923-2928.
20. Medhi RP, Chakrabarti S (2009). Traditional knowledge of NE people on conservation of wild orchids. *Ind J Traditional Knowledge*, 8: pp. 11-16.
21. Nei M, Kumar S (2000). *Evolutionary change of DNA sequences*. Molecular Evolution and Phylogenetics. Oxford University Press, NY: pp. 33-50.
22. Olmstead R, Palmer J (1994). Chloroplast DNA systematics: a review of methods and data analysis. *Am. J. Bot.* 1994, 81: pp. 1205-1224.
23. Palmer J, Jansen R, Michaels H, Chase M, Manhart J (1988). Chloroplast DNA variation and plant phylogeny. *Ann Missouri Bot Gard*, 75: pp. 1180-1206.
24. Ran JH, Wang PP, Zhao HJ, Wang XQ (2010). A test of seven candidate barcode regions from the plastome in *Picea* (Pinaceae). *J Integr Plant Biol*, 52: pp. 1109-1126.
25. Raspé O (2001). Inheritance of the chloroplast genome in *Sorbusaucuparia L* (Rosaceae). *J Hered*, 92: pp. 507-509.
26. Salazar GA, Chase MW, Arenaas MAS, Ingrouille M (2003). Phylogenetics of *Cranichideae* with emphasis on *Spiranthinae* (Orchidaceae, Orchidoideae): evidence from plastid and nuclear DNA sequences. *Am J Bot*, 90(5): pp. 777-795.
27. Sanger F, Nicklen S, Coulson AR (1977). DNA sequencing with chain-terminating inhibitors. *Proc. Natl. Acad. Sci.*, 74: pp. 5463-5467
28. Taberlet P, Coissac E, Pompanon F, Gielly L, Miquel C, Valentini A, Vermat T, Corthier G, Brochmann C, Willerslev E (2007). Power and limitations of the chloroplast *trnL* (UAA) intron for plant DNA barcoding. *Nucleic Acids Res*, 35: pp. e14.
29. Tamura K, Dudley J, Nei M, Kumar S (2007). MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *MolBiolEvol*, 24: pp. 1596-1599.
30. Valentini A, Miquel C, Taberlet P (2010). DNA barcoding for honey biodiversity. *Diversity*, 2: pp. 610-617.
31. Wolfe K, Li WH, Sharp PM (1987). Rates of nucleotide substitution vary greatly among plant mitochondrial, chloroplast, and nuclear DNAs. *Proc Natl AcadSci USA*, 84: pp. 9054-9058.
32. Xiao J-H, Wang N-X, Li Y-W, Murphy R W, Wan D-G, Niu L-M, Hu H-Y, Fu Y-G, Huang DW (2010). Molecular approaches to identify cryptic species and polymorphic species within a complex community of fig wasps. *PLoS One*, 5(11): pp. e15067
33. Yao H, Song J, Liu C, Luo K, Han J, Li Y, Pang X, Xu H, Zhu Y, Xiao P, Chen S (2010). Use of ITS2 region as the universal DNA barcode for plants and animals. *PLoS One*, 5: pp. e13102.

Corresponding Author

Komal*

Research Scholar of OPJS University, Churu, Rajasthan