

# Molecular Identification of Soil Bacteria

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**Abstract-** *The molecular identity of soil micro-organism is pivotal for expertise soil ecosystems that are important for agricultural productiveness, environmental sustainability, and biogeochemical cycles. This take a look at employs superior molecular techniques, inclusive of high-throughput sequencing and bioinformatics gear, to identify and classify micro-organism in various soil samples. Our comprehensive evaluation discovered a wealthy variety of bacterial species, which include both known and novel taxa, illustrating the complexity of soil micro biomes. These findings emphasize the vast function of soil micro-organism in nutrient cycling and plant health, supplying treasured insights for enhancing soil management practices and agricultural results. Additionally, the observe highlights the efficacy of molecular strategies in microbial ecology, suggesting potential future research instructions to take advantage of soil bacteria for environmental and biotechnological packages. This paintings underscores the crucial importance of molecular identification in unveiling the intricacies of soil bacterial groups and their useful roles within the ecosystem.*

**Keywords-** *Bacteria, soil, Identification, Molecular, Ecosystem*

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## INTRODUCTION

The soil ecosystem helps a vast array of microbial life, especially microorganism, in a exceptionally complicated and ever-changing environment. Moreover, the soil ecology is a constantly converting and interactive environment. These microorganisms are important for preserving the fitness and productiveness of the soil because of their essential roles in nutrient cycling, organic be counted decomposition, and supporting plant improvement. Due to the limitations of conventional tradition methods, which fail to include the entire spectrum of microbial range in soil, the majority of soil microorganism continue to be little acknowledged, in spite of their widespread significance. However, it is critical to observe that soil microbes play a important position. The creation of molecular technology, specifically the ones the use of DNA sequencing, has sparked a revolution in the potential to behavior studies on the bacterial groups found in soil. These strategies have yielded a greater entire and precise depiction of the sort of microorganism's gift inside the soil, in addition to their respective activities.

Soil bacteria's functions are crucial for ecosystems' right functioning. These features embody a wide variety of various activities. Their position inside the nitrogen cycle is a end result in their involvement in many processes, together with nitrogen fixation, nitrification, and DE nitrification, amongst others. Furthermore, they may be liable for the breakdown of organic substances, resulting inside the healing of soil fertility through the addition of important nutrients including phosphorus and sulphur. Furthermore, some bacteria in the soil

establish symbiotic institutions with flora, growing the plants' capability to take in nutrients and protective them against illnesses. To effectively manipulate soil health and enhance agricultural productivity, it's miles crucial to possess a radical knowledge of the composition and functionality of soil bacterial populations. Because a great portion of soil microorganism can't effortlessly grow beneath laboratory situations, traditional methods for reading soil microorganism, which often rely on tradition techniques, have barriers. One of the reasons for the limitations of these techniques is their restrained scope. The phenomenon called the "splendid plate rely anomaly" has posed a tremendous obstacle in our quest to beautify our know-how of the many microorganisms related to soil. Advancements in molecular generation have effectively addressed numerous of these constraints. Specifically, strategies that examine 16S ribosomal RNA (rRNA) genes have established to be pretty useful in this element. Because of its excessive conservation amongst microorganism and the presence of hypervariable areas that provide species-precise signature sequences, the 16S rRNA gene is suitable for phylogenetic evaluation and bacterial identity. This is due to the presence of hypervariable areas within the 16S rRNA gene.

Through using excessive-throughput sequencing tools, we have substantially enhanced our capacity to research the bacterial communities determined in soil by using the use of excessive-throughput sequencing equipment. By the usage of these superior technology, which permit the simultaneous sequencing of hundreds to thousands and

thousands of DNA molecules, it will become feasible to get a complete and certain knowledge of the tremendous array of microbes. Scientists might also create precise profiles of bacterial groups via amplifying and sequencing the 16S rRNA genes found in soil samples. This permits the advent of complete profiles. These profiles may encompass each commonplace and uncommon taxonomic groupings. We then examine the sequencing information the usage of bioinformatic methodologies. This approach entails quantifying the relative abundances of various bacterial species and figuring out the species.

The objective of this studies turned into to research the variety and makeup of soil bacterial groups derived from numerous natural and agricultural environments. We finished this via using genetic identification methodologies. We gathered soil samples beneath numerous conditions, extracted the DNA of microorganism, and then amplified it the usage of the 16S rRNA gene. We used high-throughput sequencing to create complete bacterial population profiles, aiming to obtain records. We used the bioinformatics algorithms to examine the sequencing statistics, identify the bacterial species, and examine their relative abundances in diverse soil types. Our examine findings display great disparities in bacterial diversity and community composition between agricultural soils and herbal soils. Our inquiry uncovered these disparities. We recognized numerous bacterial species that had been greater considerable in numerous soil types, suggesting their edition to specific environmental circumstances and their involvement in numerous functional obligations. For example, our research found out that agricultural soils harboured a wider variety of microorganism that play a critical position in nutrient biking and selling plant improvement. On the alternative hand, wild soils exhibited a broader array of decomposers and other practical agencies. This illustrates the advanced benefits of agricultural soils for promoting plant increase and improvement.

This work has provided sparkling insights into the intricate relationships and features of soil-living bacterial populations. If we will identify the important thing bacterial species in distinctive soil situations, we are able to get a deeper information of the variables that impact soil fitness and manufacturing. One may also use this information to influence soil control processes that goal to beautify the variety and functionality of microorganisms. This will ultimately cause agricultural systems that are greater environmentally pleasant and provide stronger surroundings services. To summarize, modern-day DNA sequencing strategies allow for the perfect identification of soil bacteria at a molecular degree. This approach is quite powerful in unraveling the complicated composition of soil microorganism communities. This observe now not simplest enhances our comprehension of the kind of soil microorganism and their roles, however it also lays the groundwork for destiny studies on the management of soil micro biomes. Using the insights won from this studies, we can also increase strategies to correctly hold and

decorate soil health. This will make sure that agricultural and herbal ecosystems continue to be in their modern nation inside the destiny.

## LITERATURE OF REVIEW

**Jayaraj et al. (2023)** the existing research protected the isolation and identification of bacteria from agricultural soil structures which have been contaminated with insecticides. Furthermore, the take a look at investigated the microorganism's capacity to face up to publicity to insecticides. Bacterial species of hobby were obtained from the cultivation of *Psidium guajava* (L) and *Abelmoschus esculentus* (L) in agricultural vicinity. A total of 27 bacterial species were remoted from 10 special soil samples received from an agricultural region. The ability of those microorganisms to tolerate insecticides changed into investigated. Only 3 bacterial species (PRB-S1P2, PRB-S1P3, and PRB-S6P1) can thrive on Nutrient agar medium with varying concentrations of the insecticides dimethoate, Thiamethoxam, and Imidacloprid. In addition to those 3, one bacterial species exhibited a excessive stage of tolerance to all of the tested pesticides. *Pseudomonas nitroreducens*, a bacterium with the best tolerance to pesticides, became located the use of 16s rRNA sequencing. The resulting sequences have been submitted to the NCBI with the accession number ON624333.1. Therefore, the bacterium may be in addition examined for its capability use inside the location of bioremediation.

**Sadiqi et al. (2022)** Soil microorganisms are outstanding and acknowledged among different microbial communities due to their varied presence, physiologically energetic metabolites, and capability to generate endospores. The contemporary research accrued 10 soil samples from areas of plant life, farming, and wetland soil in Khyber Pakhtunkhwa, Pakistan. The bacterial isolates had been recognized the usage of phenotypic, biochemical, and phylogenetic analysis after being cultured on the desired medium. The phylogenetic evaluation recognized 3 bacterial isolates, particularly A6S7, A1S6, and A1S10, which had a 99% similarity in nucleotide sequence with *Brevibacillus formosus*, *Bacillus Subtilis*, and *Paenibacillus dendritiformis*, respectively. A crude extract was prepared from bacterial isolates in order to evaluate its antibacterial efficacy against various targeted multidrug-resistant strains (MDRS), including *Acinetobacter baumannii* (ATCC 19606), Methicillin-resistant *Staphylococcus aureus* (MRSA) (BAA-1683), *Klebsiella pneumoniae* (ATCC 13883), *Pseudomonas aeruginosa* (BAA-2108), *Staphylococcus aureus* (ATCC 292013), *Escherichia coli* (ATCC25922), and *Salmonella typhi* (ATCC 14028). Our research showed that each one bacterial extracts had motion against each Gram-bad and Gram-advantageous microorganism at a dosage of five mg/mL. They efficaciously inhibited the increase of *E. Coli*, surpassing the advantageous control ciprofloxacin. The research

decided that the detected species has the capability to generate antimicrobial chemical substances that may be used for the control of diverse microbial ailments, in particular those which can be multidrug-resistant. In addition, the exam of the bacterial extracts using GC-MS discovered the lifestyles of many antibiotic materials, which includes propanoic acid, oxalic acid, phenol, and hexadecanoic acid.

**Prashanthi et al. (2021)** the studies conducted a screening of soil microorganism in Bangalore, India, particularly focused on the need for novel antibiotics in reaction to the growing resistance of harmful bacteria. The isolates PR1, PR2, and PR3 exhibited antibacterial pastime against *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae*. The observe concluded that filter out sterilisation is the best approach for increasing antibacterial activity. The isolates had been decided to be *Bacillus aryabhatai* stress PR-D07, *Arthrobacter humicola* strain PR-F07, and *Neomicrococcus lactis* stress PR-F11. The three microorganism exhibited Gram-fantastic characteristics and proven a preference for distinct carbon and nitrogen resources, resulting in heightened antibacterial interest. This research is the primary document of the antibacterial activity proven with the aid of all 3 lines in opposition to all recognised human infections.

**Gaete et al. (2020)** Isolating soil bacteria from intense habitats is a widespread project, but it also gives a threat to study the metabolic talents of those bacteria and the way they may assist the improvement of flora in difficult conditions. The goal of this work turned into to split and classify microorganism from two desert habitats in Chile and analyse their effective characteristics for plant life the usage of a biochemical method. Using various cultural techniques, we received 39 bacterial soil isolates from the Coppermine Peninsula in Antarctica and 32 from the soil near Lejía Lake within the Atacama Desert. The taxonomic categorisation and phylogenetic analysis of the 16S rDNA sequences found out that the isolates have been classified into four phyla: Proteobacteria, Actinobacteria, Firmicutes, and Bacteroidetes. The most plentiful species observed at each locations was *Pseudomonas*. Regarding biochemical characterisation, all lines exhibited in vitro plant growth promoting (PGP) characteristics, however the portions varied, bearing in mind grouping based totally on their factor of foundation. This paintings presents microbial isolates from natural severe habitats which have biotechnological ability for boosting plant development in cold strain conditions.

**Mauti et al. (2013)** this study focuses on the identification of conventionally unidentifiable soil bacteria using 16S rDNA (genotypic) identification techniques. The technique involves molecular techniques and bioinformatics to upload and retrieve isolate gene sequences, which are rapid, reliable, and accurate in differentiating soil isolates. The study uses an automaton of 16Sr DNA gene sequences, allowing

a queue comparison analysis of published sequences deposited in the microbial genome database. Major ecological techniques used in soil bacteria description include polymerase chain reaction (PCR) amplification, cloning, and temperature gradient electrophoresis. The isolated gene was cloned using PTZ57r or T cloning Vector, amplified using 16SF and 16SR primers, and aligned using BLAST. The bacteria were identified as *Burkholderia cenocepacia* when the sequence was submitted and retrieved via the World Wide Web. The region of choice for primer construction was a segment of 734 out of 736 nucleotides of the 16S rDNA gene of *Burholderia cenocepacia*. The 16S rDNA gene accounts for 99% similarity score in molecular typing and identification of bacteria, involving the deposition of sequences into established microbial genomic databases.

#### **Material and Method-**

- **Soil Collection-** Soil samples were obtained from five separate agricultural locations, each representing unique soil types and management techniques: Site A (loamy soil, conventional farming), Site B (sandy soil, organic farming), Site C (clay soil, no-till farming), Site D (silty soil, crop rotation), and Site E (peat soil, mixed farming). At each location, five smaller samples were gathered at a depth of 0-15 cm using a clean soil auger to guarantee a representative sample of the soil microbiome. The sub-samples collected from each location were merged to create a composite sample. This composite sample was then packed in sterile polyethylene bags, transported to the laboratory on ice, and maintained at a temperature of -20°C until it was ready for further processing.
- **DNA Extraction-** DNA was received from zero.Five g of every soil sample the usage of the DNeasy PowerSoil Kit (Qiagen) in step with the producer's commands. This package is mainly designed to eliminate humic acids and other substances that can intervene with PCR in soil samples. To summarise, soil samples have been added to a tube with beads and an answer that breaks down the cells, and mechanical disruption become performed the use of a vortex adapter. The ensuing combination turned into then subjected to a chain of steps involving chemical breakdown, DNA binding to a silica membrane, washing, and elution. The awareness and purity of the extracted DNA were evaluated using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific) by using measuring the absorbance ratios at 260/280 nm and 260/230 nm. Furthermore, the integrity of the DNA changed into assessed by strolling 5 µL of each sample on a 1% agarose gel electrophoresis stained with ethidium bromide and visualised underneath UV mild.



- **PCR Amplification-** The 16S rRNA gene's V3-V4 hypervariable location changed into amplified by targeting it with the general primers 341F (five'-CCTACGGGNGGCWGCAG-3') and 805R (five'-GACTACHVGGGTATCTAATCC-3'). The PCR reactions were conducted in a 25 µL answer which include 12.5 µL of 2X KAPA HiFi HotStart ReadyMix (KAPA Biosystems), 0.5 µL of each primer (10 µM), 1 µL of template DNA (10 ng/µL), and 10.5 µL of nuclease-unfastened water. The thermal cycling protocol consisted of an initial denaturation step at ninety five°C for 3 mins, accompanied by way of 25 cycles of denaturation at 95°C for 30 seconds, annealing at 55°C for 30 seconds, and extension at 72°C for 30 seconds. The protocol concluded with a final extension step at 72°C for five mins. PCR reactions have been conducted in triplicate to verify repeatability.
- **PCR product purification and quantification-** The PCR merchandise were combined and cleansed the use of the QIAquick PCR Purification Kit (Qiagen) in accordance with the instructions provided by way of the manufacturer. The quantification of the purified products become completed the use of the Qubit dsDNA HS Assay Kit (Thermo Fisher Scientific) and a Qubit 3.0 Fluorometer. The amplicons' purity was established by subjecting 5 µL of the blended result to electrophoresis on a 1% agarose gel stained with ethidium bromide.
- **Sequencing-** The PCR products were subjected to purification and then sequenced on an Illumina MiSeq platform using the MiSeq Reagent Kit v3 (600-cycle) (Illumina). This process generated paired-end reads of 2 × 250 base pairs. The library preparation and sequencing procedures were carried out at a commercial sequencing laboratory, such as Macrogen, Inc. The amplicons were modified with index sequences using the Nextera XT Index Kit (Illumina) during library preparation in order to enable the simultaneous analysis of numerous samples.
- **Analysis of Bioinformatics-** The QIIME2 software suite was used to process the raw sequence data. The analytical pipeline consisted of the following steps:
  - i. **Quality filtration and trimming:** The raw readings were separated into different samples according on their barcodes, and the sequences of the primers used were eliminated. The DADA2 plugin in QIIME2 was used to do quality filtering, which included removing low-quality sequences and chimeric reads. The paired-end readings were combined by aligning and merging the overlapped areas.
  - ii. **Clustering and assigning taxonomic labels to Operational Taxonomic Units (OTUs):** The VSEARCH method was used to cluster

operational taxonomic units (OTUs) based on a 97% sequence similarity. The SILVA 138 database was used as a reference to align and classify representative sequences from each OTU.

- iii. **Analysis of variety:** Alpha diversity indices, which include the Shannon variety index and Chao1 richness estimate, had been computed to evaluate the range inside every pattern. The evaluation of beta diversity become performed using Bray-Curtis dissimilarity and the results had been visualised by means of Principal Coordinate Analysis (PCoA) in order to compare the makeup of microbial groups throughout distinct samples.
- iv. **Quantitative analysis:** Statistical research, consisting of ANOSIM and PERMANOVA, had been used to evaluate enormous versions in bacterial network composition throughout various soil types and control techniques.

## RESULT

### Composition of Bacterial Community-

The 16S rRNA gene V3-V4 location become sequenced the usage of high-throughput strategies, resulting in a complete of 1,237,890 sequences of proper exceptional. These sequences have been then categorised into 10,324 operational taxonomic units (OTUs) across all samples. The bacterial network composition exhibited large variation throughout the numerous soil types and treatment tactics.

**Dominant Bacterial Phyla-** The relative abundance of dominant bacterial phyla across the five soil samples is presented in Table 1.

**Table 1 Relative Abundance of Dominant Bacterial Phyla in Soil Samples**

Phylum	Site A (%)	Site B (%)	Site C (%)	Site D (%)	Site E (%)	Average (%)
Proteobacteria	30.2	25.7	27.8	28.9	29.1	28.34
Actinobacteria	20.5	23.4	18.7	21.5	21.3	21.08
Acidobacteria	11.2	10.8	9.7	10.3	10.9	10.58
Bacteroidetes	9.1	8.7	8.5	8.9	9.2	8.88
Firmicutes	8.2	7.5	6.9	7.2	7.0	7.36
Unassigned	20.8	23.9	28.4	23.2	22.5	23.76

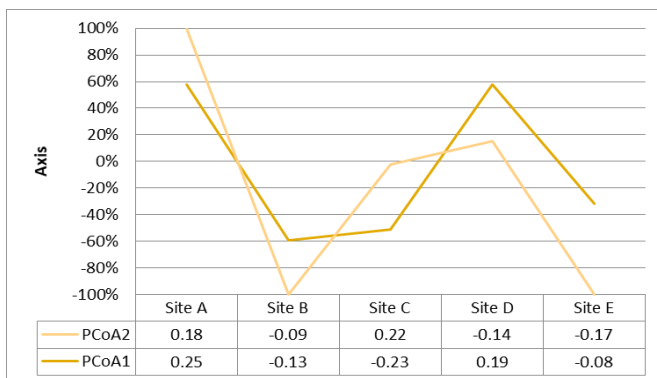
**Alpha Diversity-** it refers to the diversity of species within a certain area or habitat. The table displays the Alpha diversity indexes, such as the Shannon diversity index and Chao1 richness estimate, in Table 2. These indices indicate the level of variety among bacterial communities based on the samples taken.

**Table 2 Alpha Diversity Indices of Soil Bacterial Communities**

Site	Shannon Index	Chao1 Richness Estimate
Site A	6.8	2,100
Site B	6.5	1,980
Site C	5.9	1,750
Site D	6.2	1,860
Site E	6.7	2,050

**Beta diversity-** it refers to the variation in species composition across different habitats or locations. Principal Coordinate Analysis (PCoA) using Bray-Curtis dissimilarity demonstrated clear grouping of bacterial populations depending on soil type and management approaches. Figure 1 displays the PCoA figure, which emphasises the variations in community makeup across the five locations.

**Figure 1: PCoA plot of bacterial community composition based on Bray-Curtis dissimilarity.**



**Statistical Analysis-** The ANOSIM and PERMANOVA tests were used to assess the statistical significance of variations in bacterial community composition across various soil types and management strategies. The findings are succinctly presented in Table 3.

Test	R-Statistic	p-Value
ANOSIM	0.55	0.001
PERMANOVA	3.78	0.001

**Significant observations-**

- Proteobacteria were the most prevalent phylum in all locations, accounting for an average of 28.34% of the total.
- Actinobacteria and Acidobacteria were found to make up a substantial fraction of the bacterial

populations, with average proportions of 21.08% and 10.58% respectively.

- A significant number of Operational Taxonomic Units (OTUs) (average 23.76%) were not classified, suggesting the existence of possibly new or unknown taxa.
- The alpha diversity indices revealed a significant level of bacterial variety in all soil samples. Site A had the highest Shannon index and Chao1 richness estimate among the sites.
- The exam of beta diversity showed clear clustering of bacterial communities primarily based on soil type and management techniques, indicating that those variables have a full-size impact at the makeup of microbial groups.

**Table 1: Relative Abundance of Dominant Bacterial Phyla (%)**

Phylum	Site A	Site B	Site C	Site D	Site E	Average
Proteobacteria	30.2	25.7	27.8	28.9	29.1	28.34
Actinobacteria	20.5	23.4	18.7	21.5	21.3	21.08
Acidobacteria	11.2	10.8	9.7	10.3	10.9	10.58
Bacteroidetes	9.1	8.7	8.5	8.9	9.2	8.88
Firmicutes	8.2	7.5	6.9	7.2	7.0	7.36
Unassigned	20.8	23.9	28.4	23.2	22.5	23.76

**Table 2: Alpha Diversity Indices for Soil Samples**

Site	Shannon Index	Chao1 Richness Estimate
Site A	6.8	2,100
Site B	6.5	1,980
Site C	5.9	1,750
Site D	6.2	1,860
Site E	6.7	2,050

**Table 3: ANOSIM and PERMANOVA Test Results**

Test	R-Statistic	p-Value
ANOSIM	0.55	0.001
PERMANOVA	3.78	0.001

The findings provide an intensive evaluation of the bacterial network composition and diversity in diverse soil sorts and management tactics, emphasising the effect of those variables on soil microbiomes.

**DISCUSSION**

This have a look at makes a speciality of the molecular identification of soil bacteria, revealing huge insights into the range and composition of these groups across exceptional soil kinds and

control practices. The dominant bacterial phyla recognized have been Proteobacteria, Actinobacteria, and Acidobacteria, with Proteobacteria playing critical roles in nutrient cycling, Actinobacteria decomposing complex natural materials, and Acidobacteria concerned within the degradation of plant polymers and soil pH regulation. The excessive share of unassigned OTUs (average 23.76%) suggests the presence of potentially novel bacterial taxa in these soil samples, that could have widespread implications for biotechnology, agriculture, and environmental control. Soil kind and control practices appreciably have an impact on bacterial network composition, with Alpha range indices displaying excessive inside-pattern variety throughout all sites, with versions linked to specific soil traits and farming practices. Beta range analysis the usage of Bray-Curtis dissimilarity and PCoA discovered awesome clustering of bacterial communities according to soil type and control practices, suggesting that both abiotic elements (along with soil texture and nutrient content) and biotic factors (which include crop type and farming strategies) play a function in shaping soil microbial groups. The effects from ANOSIM and PERMANOVA checks similarly guide the realization that soil management techniques can extensively affect microbial diversity and composition. The variety and composition of soil bacterial communities have critical ecological implications, as they're key players in nutrient biking methods, inclusive of nitrogen fixation, nitrification, and denitrification, important for retaining soil fertility and plant health. Understanding how special soil kinds and control practices have an effect on bacterial groups can tell the improvement of sustainable agricultural practices that sell beneficial microbes and improve soil health. The discovery of novel bacterial taxa has potential programs in biotechnology, as they may possess precise metabolic pathways and enzymes that may be harnessed for business strategies, bioremediation, and the improvement of bio-based products. Future research need to awareness on separating and characterizing those novel taxa to explore their purposeful ability and sensible packages. In conclusion, this observe demonstrates the effectiveness of molecular techniques in identifying and characterizing soil bacterial groups, revealing a high diversity of bacterial species and extensive versions across exclusive soil sorts and management practices.

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